

前瞻与评论

基于文献统计学的我国新疆特殊环境微生物新种资源挖掘概况及展望

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摘要: 【背景】我国新疆地域辽阔, 气候类型多样, 具有多样性极高的特有微生物资源。【目的】基于文献统计学, 探究新疆特殊环境微生物新种资源挖掘现状与趋势。【方法】以“Xinjiang”为关键词, 在 IJSEM 等发表微生物新种的主流刊物上检索并筛选出 2000–2021 年 22 年间源自新疆地区的新种文献合计 321 篇共计 348 株新种用于本研究计量分析, 使用 VOS viewer 对其关键词、发表时间、作者及单位进行可视分析。【结果】新种文献发表的数量整体呈现波动上升趋势, 主要发文单位是云南大学和塔里木大学, 高频关键词为土壤微生物, 对全部新种的种属分类、分离获得的方法、潜在的应用价值进行归纳总结。【结论】根据文献统计学(bibliometrics)分析了新疆地区孕育的微生物新种资源概况及相关研究趋势, 为将来系统性地开发新疆地区特有微生物种质资源提供参考。

关键词: 新疆; 新种鉴定; 微生物新种资源; 可视化

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Overview and prospect of newly discovered microbial resources in the special environments of Xinjiang in China based on bibliometrics

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Abstract: [Background] Xinjiang of China, with its vast land and diverse climate types, which harbors high diversity of endemic microbial resources. [Objective] Based on bibliometrics, to explore the status and trend of newly discovered microbial resources in the special environments of Xinjiang. [Methods] We searched against IJSEM and other mainstream journals that publish newly discovered species with “Xinjiang” as the keyword and the time interval of 2000 to 2021. A total of 321 articles involving 348 strains of endemic species newly discovered in Xinjiang were screened out and used for bibliometric analysis. VOS viewer was used for the visual analysis of the keywords, publishing dates, authors and affiliations. [Results] The number of literatures published on new species showed an overall trend of fluctuation and increase, and the major publishing affiliations were Yunnan University and Tarim University. The high-frequency keyword was soil microorganisms, the taxonomic status, isolation methods, and potential application values of all the newly discovered species were summarized. [Conclusion] Based on bibliometrics, the overview and related research trend of new microbial resources in Xinjiang were analyzed. We hope that this paper will provide a reference for the systematic mining of microbial germplasm resources endemic to Xinjiang in the future.

Keywords: Xinjiang; identification of new species; new microbial resources; visualization

新疆位于欧亚大陆中心，我国西北边陲，面积 166 万 km²，占全国陆地面积的 1/6^[1]。四周远离海洋，被高原、高山环绕，属温带大陆性气候，冬季漫长严寒，夏季炎热干燥，春秋季短促且变化剧烈^[2]。在地形、气候、土壤类型等各种自然因素的共同作用下，新疆具有我国最干、最热、最冷、温差最大、海拔最低的沙漠、戈壁、冰川、盐湖等多种极端环境和自然生态系统^[3]。适应于这些自然生态环境的微生物在长期进化过程中形成了丰富多样的特有

微生物类群，具有极大的研究和开发价值。李泉泉等^[4]对新疆七角井盐湖和南湖碱湖可培养极端嗜盐菌组成及多样性进行研究，筛选到多株耐高盐环境的产酶功能菌；田蕾等^[5]调查艾丁盐湖沉积物细菌多样性时，挖掘到了新分类的菌属；李娟等^[6]对塔克拉玛干沙漠微生物的组成及特性进行描述，经统计有 17 株新种被正式发表。截至目前，这些盐碱含量高、紫外辐射强、营养物质贫瘠等特殊环境因素长期形成非生物胁迫，导致当地微生物类群具有独特的

基因类型、特殊的生理机制及代谢产物。本文从新疆特殊生境微生物新种的分离源、分离方法、潜在的应用价值进行统计分析，并对今后系统开发新疆地区微生物新种资源提出展望，以期为后续的研究提供参考。

1 材料与方法

1.1 数据检索和分析条目

本文所有文献来源于 Web of Science 数据库中 *International Journal of Systematic and Evolutionary Microbiology*、*Antonie van Leeuwenhoek*、*Current Microbiology*、*Archives of Microbiology* 和 *The Journal of Microbiology*。检索年限限定为 2000 年 1 月 1 日–2021 年 12 月 31 日，设定的检索词为“Xinjiang”。

1.2 分析软件及方法

经检索符合条件的所有文献导入 Endnote (Version X9)，通过分析文献数据后导出使用 OriginPro 8.5 和 VOS viewer 分析，数据整理统计采用 OriginPro 8.5，图像可视化采用 VOS viewer。

2 结果与分析

2.1 新疆地区微生物新种文献发表分析

2.1.1 新种文献发表信息概述

以“Xinjiang”为关键词，在 IJSEM 等刊物检索出 634 篇文献。为保证数据的准确性，以检索词“Xinjiang”出现在新种菌株分离地作为二次筛选条件，对检索出的文献进行了去重、删除不符合要求的文献，筛选整理后得到 2000–2021 年间发表的新种文献合计 321 篇。

新种文献发表的数量整体呈现波动上升趋势。2000–2007 年每年发文数量不超过 5 篇，自 2008 年高盐环境分离培养基出现^[7]后发文数量大幅增加，其中最大值出现在 2015 年，新种

文献发表数量为 32 篇(图 1A)。

2.1.2 研究作者及单位机构分析

由于各研究机构通过合作方式发表新种的文献比较多，主要参考各文献第一完成单位进行分类汇总，统计各单位的发文数量，用以评价新疆地区新种资源研究机构的分布与贡献度。整理出的 321 篇论文来自 202 名作者和 54 个单位，其中中国云南大学和塔里木大学是主要的发文单位，李文均和张利莉是主要通讯作者，我国新疆单位与非新疆单位分别占 28.97% 和 71.03% (图 1B、1C)，非新疆单位发表新种数量明显多于我国新疆单位，国内其他研究机构在我国新疆地区环境的微生物资源挖掘方面发挥主导作用。

2.1.3 高频关键词聚类分析

将待分析的文献由 Endnote 转入 VOS viewer，出现 5 次以上(剔除细菌鉴定方面术语)关键词制作标签视图，利用 VOS viewer 构建关键词共现网络(图 2)。对新疆地区新种文献关键词频次统计，土壤、湖泊、沙漠、沉积物、冰山、根际、植物内生环境出现频次由高至低；放线菌、链霉菌、盐单胞菌、芽孢杆菌细菌出现频次由高至低。其中氯化钠代谢与湖泊关联最高，其次是土壤，再者是放线菌，这与分离源多为盐湖及盐土环境一致。柽柳根际土壤是被较多研究的，胡杨内生菌新种是被发表最频繁的。总体表明该地区特有生境微生物资源的多样性及独特性。

目前从新疆盐湖分离的嗜盐、耐盐微生物新种有 60 株，主要源于七角井盐湖、艾丁湖、硝尔库勒盐湖、艾比湖、阿牙克库木湖等盐湖；在研究荒漠生态系统可培养细菌时，分离自塔克拉玛干沙漠、古尔班通古特沙漠、吐鲁番和库尔勒等地区荒漠环境新种共 40 株，其干旱、寡营养和极强的太阳辐射等条件造就了沙漠中

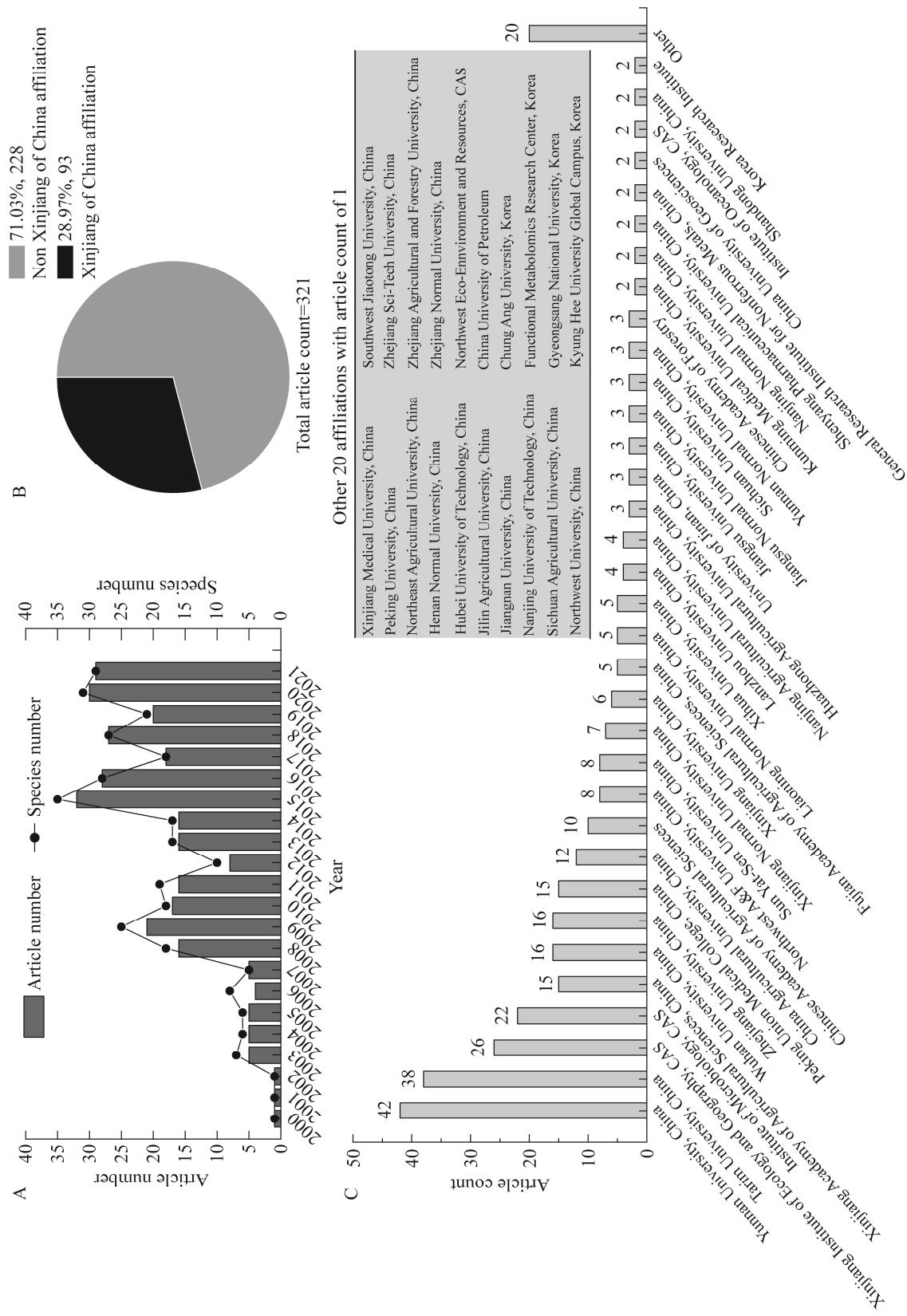


图 1 2000—2021年之间已发表的源自我国内新疆地区的新型文章情况概览
Figure 1 Overview of new species published from Xinjiang of China between 2000 and 2021. A: The proportion of affilations. B: The distribution of published affiliations.

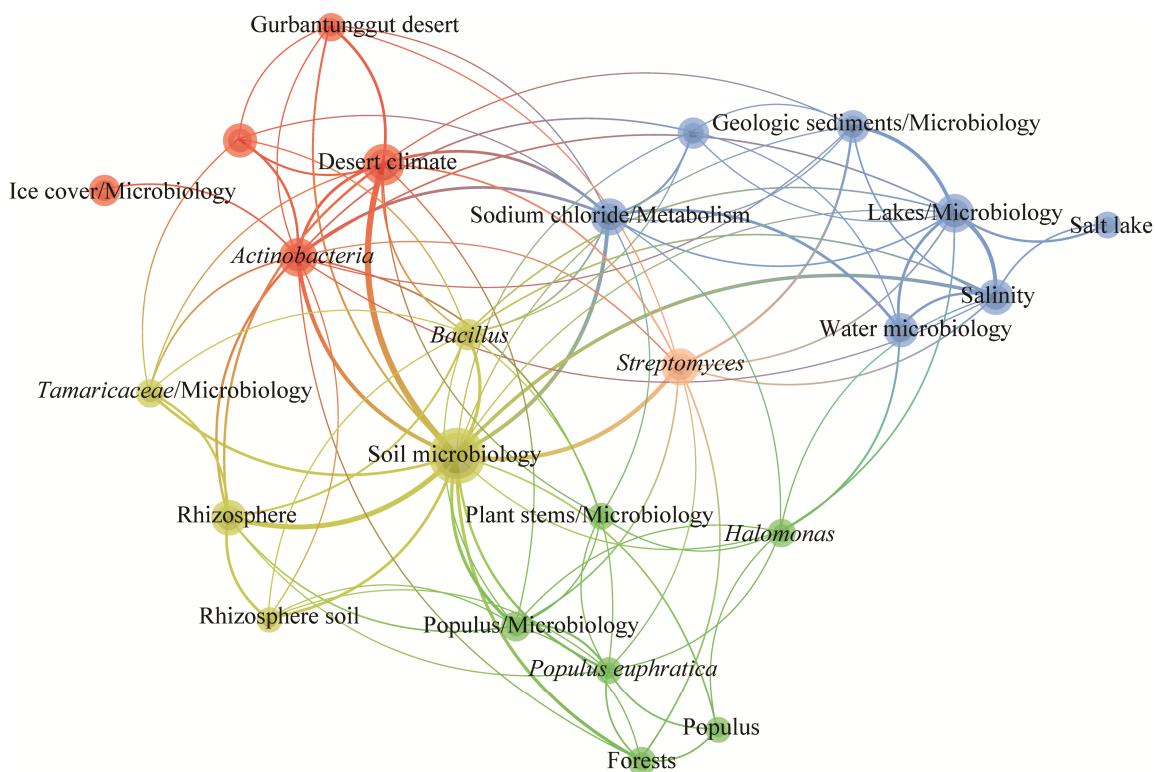


图 2 由 VOS viewer 呈现出的新种文章高频关键词分析

Figure 2 High frequency keyword analysis of new species articles presented by VOS viewer.

细菌耐旱、耐高温、耐辐射的特性；此外，由于新疆地区的冰川环境常年覆盖冰雪，处于低温状态，孕育了天然的嗜/耐低温菌种资源库。近年分离自天山 1 号冰川等环境的嗜冷/耐冷新种共 21 株，主要为冷杆菌属(*Cryobacterium*)^[8-13]、黄杆菌属(*Flavobacterium*)^[14-19]；其余分离源还包括胡杨^[20-21]、柽柳^[22-23]、骆驼刺^[24-25]、梭梭^[26-27]、银砂槐^[28]等荒漠植物及一些新疆特有极端环境，如罗布泊^[29-30]、楼兰古城^[31-32]、盐矿^[33-34]、受辐射的土壤等^[35]。

2.2 微生物新种分类地位分布

2000–2021 年间，采自新疆地区各类环境样品中分离并已发表的新种 348 株(文章共计 321 篇)。所有新种细菌域 331 株，其中放线菌门(*Actinomycetota*) 170 株，分属于链霉菌属(*Streptomyces*) 29 株、糖霉菌属(*Glycomyces*)

9 株、拟诺卡氏菌属(*Nocardiopsis*) 11 株等；假单胞菌门(*Pseudomonadota*) 62 株，分属于盐单胞菌属(*Halomonas*) 10 株、假单胞菌属(*Pseudomonas*) 6 株、根瘤菌属(*Rhizobium*) 5 株等；芽孢杆菌门(*Bacillota*) 51 株，分属于芽孢杆菌属(*Bacillus*) 11 株、类芽孢杆菌属(*Paenibacillus*) 8 株、大洋芽孢杆菌属(*Oceanobacillus*) 6 株等；拟杆菌门(*Bacteroidota*) 44 株，分属于海洋杆菌属(*Pontibacter*) 9 株、黄杆菌属(*Flavobacterium*) 8 株、鞘氨醇杆菌属(*Sphingobacterium*) 5 株等；异常球菌门(*Deinococcota*) 4 株，属于奇球菌属(*Deinococcus*)。古菌域 15 株，其中广古菌门(*Euryarchaeota*) 15 株，分别隶属于钠线菌属(*Natrinema*)、盐陆生菌属(*Haloterrigena*)、盐惰菌属(*Halopiger*)、纳杆菌属(*Natribaculum*)、嗜盐碱红菌

(*Natronorubrum*)等 11 个属。真核生物域 2 株, 分别采自小蠹虫和野生植物叶片, 隶属于子囊菌门(*Ascomycota*)假丝酵母菌属^[36] (*Candida*) 和担子菌门(*Basidiomycota*) *Udeniomyces*^[37]。其中归属于放线菌门的菌属数量最多, 原因可能与分离环境多为土壤以及开展新疆极端环境下放线菌资源生物多样性研究有关; 同时放线菌能产生大量具有生物活性的次级代谢产物, 是发现新药物的重要资源; 由于链霉菌属能够产生耐热和抗干燥的孢子, 可以在不利环境下长期存活, 具有较高的环境抗逆性^[38], 因此分离于沙漠、盐湖等环境的菌属中, 链霉菌属丰度占比最高, 成为该地区特殊环境中被大量发掘的微生物菌属(图 3)。

经新菌种分类地位统计分析可知, 放线菌门 3 纲 16 目 29 科 67 属, 拟杆菌门 6 纲 6 目

9 科 21 属, 假单胞菌门 3 纲 14 目 23 科 33 属, 芽孢杆菌门 2 纲 3 目 5 科 21 属, 异常球菌门 1 纲 1 目 1 科 1 属, 广古菌门 1 纲 3 目 4 科 12 属, 子囊菌门和担子菌门各 1 属(表 1)。从发表速度来看, 在 2008 年之前, 获得常见菌属较为广泛。主要为假单胞菌门的根瘤菌属(*Rhizobium*); 芽孢杆菌门的嗜盐芽孢杆菌属(*Halobacillus*)、慢生芽孢杆菌属(*Lentibacillus*)、细纤芽孢杆菌属(*Tenuibacillus*)、芽孢杆菌属(*Bacillus*); 拟杆菌门的黄杆菌属(*Flavobacterium*)、海洋杆菌属(*Pontibacter*)、薄层杆菌属(*Hymenobacter*); 放线菌门的链霉菌属(*Streptomyces*)、拟诺卡氏菌属(*Nocardiopsis*); 广古菌门的盐陆生菌属(*Haloterrigena*)、钠线菌属(*Natrinema*)。随着菌种资源的大量获得, 陆续出现了特殊菌属, 具有嗜盐特性的放线菌 *Haloechinothrix*、

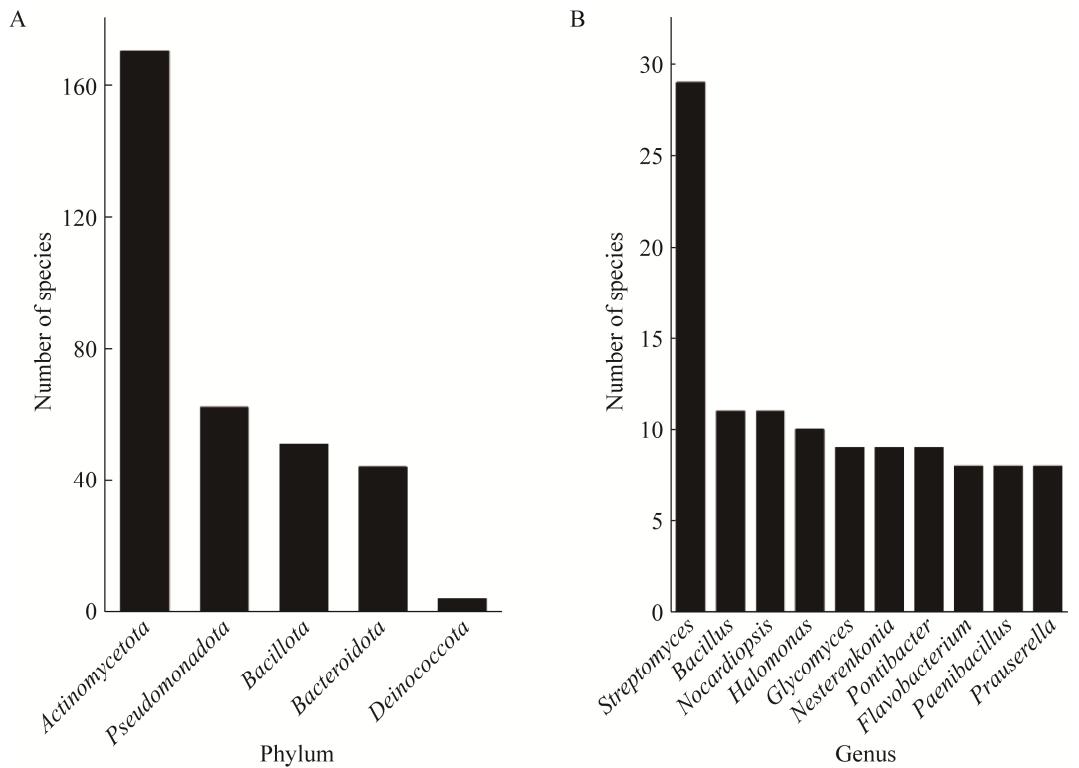


图 3 2000–2021 年间被描述的新种数量最多的细菌属和门

Figure 3 Bacterial genera and phyla with the highest number of new species described between 2000–2021.

表 1 已发表新疆地区微生物新种分类情况(2000–2021)

Table 1 Classification of published novel species in Xinjiang (2000–2021)

Phylum	Class	Order	Family	Genus	Genus number
<i>Actinomycetota</i>	<i>Actinomycetia</i>	<i>Streptomycetales</i>	<i>Streptomycetaceae</i>	<i>Streptomyces</i>	29
		<i>Streptosporangiales</i>	<i>Nocardiopsaceae</i>	<i>Nocardiopsis</i>	11
				<i>Streptomonospora</i>	5
				<i>Marinactinospora</i>	1
				<i>Lipingzhangella</i>	1
				<i>Haloactinospora</i>	1
				<i>Allosalinactinospora</i>	1
			<i>Thermomonosporaceae</i>	<i>Actinomadura</i>	1
	<i>Micrococcales</i>	<i>Micrococcaceae</i>		<i>Nesterenkonia</i>	9
				<i>Arthrobacter</i>	5
				<i>Kocuria</i>	3
				<i>Yania</i>	1
				<i>Zhihengluiella</i>	1
		<i>Microbacteriaceae</i>		<i>Cryobacterium</i>	7
				<i>Microbacterium</i>	3
				<i>Labedella</i>	3
				<i>Salinibacterium</i>	1
				<i>Pseudoclavibacter</i>	1
				<i>Planctomonas</i>	1
				<i>Mycetocola</i>	1
				<i>Frigoribacterium</i>	1
				<i>Leifsonia</i>	1
				<i>Okibacterium</i>	1
		<i>Ruaniaceae</i>		<i>Ruania</i>	1
				<i>Haloactinobacterium</i>	1
		<i>Brevibacteriaceae</i>		<i>Brevibacterium</i>	2
				<i>Bogoriellaceae</i>	1
				<i>Georgenia</i>	1
				<i>Cellulomonadaceae</i>	1
				<i>Actinotalea</i>	1
		<i>Jonesiaceae</i>		<i>Jonesia</i>	1
				<i>Intrasporangiaceae</i>	1
				<i>Ornithinicoccus</i>	1
				<i>Ornithinimicrobiaceae</i>	1
				<i>Ornithinimicrobium</i>	1
		<i>Promicromonosporaceae</i>		<i>Myceligerans</i>	1
				<i>Sanguibacteraceae</i>	1
	<i>Pseudonocardiales</i>	<i>Pseudonocardiaceae</i>		<i>Sanguibacter</i>	1
				<i>Prauserella</i>	8
				<i>Saccharopolyspora</i>	4
				<i>Saccharothrix</i>	3
				<i>Saccharomonospora</i>	3
				<i>Amycolatopsis</i>	3

(待续)

(续表 1)

Phylum	Class	Order	Family	Genus	Genus number
				<i>Haloechinothrix</i>	2
				<i>Alloactinosynnema</i>	1
				<i>Actinophytocola</i>	1
				<i>Actinokineospora</i>	1
				<i>Actinobispora</i>	1
				<i>Longimycelium</i>	1
				<i>Lechevalieria</i>	1
				<i>Yuhushiella</i>	1
				<i>Herbihabitans</i>	1
	<i>Glycomycetales</i>	<i>Glycomycetaceae</i>		<i>Glycomyces</i>	9
				<i>Haloglycomyces</i>	1
				<i>Paraglycomyces</i>	1
	<i>Propionibacteriales</i>	<i>Nocardioidaceae</i>		<i>Nocardiooides</i>	4
				<i>Aeromicrobium</i>	2
		<i>Propionibacteriaceae</i>		<i>Desertihabitans</i>	1
	<i>Corynebacteriales</i>	<i>Mycobacteriaceae</i>		<i>Hoyosella</i>	1
				<i>Mycolicibacterium</i>	1
		<i>Corynebacteriaceae</i>		<i>Corynebacterium</i>	1
	<i>Jiangellales</i>	<i>Jiangellaceae</i>		<i>Phytoactinopolyspora</i>	4
				<i>Haloactinopolyspora</i>	2
	<i>Micromonosporales</i>	<i>Micromonosporaceae</i>		<i>Actinoplanes</i>	3
				<i>Allorhizocola</i>	1
	<i>Actinopolysporales</i>	<i>Actinopolysporaceae</i>		<i>Actinopolyspora</i>	4
	<i>Geodermatophilales</i>	<i>Geodermatophilaceae</i>		<i>Blastococcus</i>	1
	<i>Kineosporiales</i>	<i>Kineosporiaceae</i>		<i>Kineococcus</i>	1
	<i>Nakamurellales</i>	<i>Nakamurellaceae</i>		<i>Nakamurella</i>	1
	<i>Nitriliruptoria</i>	<i>Egicoccales</i>	<i>Egicoccaceae</i>	<i>Egicoccus</i>	1
		<i>Egibacteriales</i>	<i>Egibacteraceae</i>	<i>Egibacter</i>	1
	<i>Acidimicrobia</i>	<i>Acidimicrobiales</i>	<i>Ilumatobacteraceae</i>	<i>Desertimonas</i>	1
<i>Pseudomonadota</i>	<i>Alphaproteobacteria</i>	<i>Hyphomicrobiales</i>	<i>Rhizobiaceae</i>	<i>Rhizobium</i>	5
				<i>Sinorhizobium</i>	1
			<i>Phyllobacteriaceae</i>	<i>Chelativorans</i>	2
				<i>Mesorhizobium</i>	5
			<i>Aurantimonadaceae</i>	<i>Aurantimonas</i>	1
			<i>Devosiaceae</i>	<i>Pelagibacterium</i>	1
			<i>Brucellaceae</i>	<i>Ochrobactrum</i>	1
			<i>Hyphomicrobiaceae</i>	<i>Limoniibacter</i>	1
	<i>Rhodobacterales</i>	<i>Rhodobacteraceae</i>		<i>Paracoccus</i>	2
				<i>Frigidibacter</i>	2

(待续)

(续表 1)

Phylum	Class	Order	Family	Genus	Genus number
				<i>Falsirhodobacter</i>	1
				<i>Defluviimonas</i>	1
		<i>Rhodospirillales</i>	<i>Azospirillaceae</i>	<i>Arenibaculum</i>	1
				<i>Desertibacter</i>	2
				<i>Skermanella</i>	3
			<i>Acetobacteraceae</i>	<i>Sabulicella</i>	1
		<i>Sphingomonadales</i>	<i>Erythrobacteraceae</i>	<i>Altererythrobacter</i>	1
				<i>Croceibacterium</i>	1
			<i>Sphingomonadaceae</i>	<i>Novosphingobium</i>	1
		<i>Rhizobiales</i>	<i>Methylocystaceae</i>	<i>Flaviflagellibacter</i>	1
		<i>Parvularculales</i>	<i>Parvularculaceae</i>	<i>Amphiplicatus</i>	1
<i>Betaproteobacteria</i>	<i>Burkholderiales</i>		<i>Alcaligenaceae</i>	<i>Alcaligenes</i>	1
			<i>Comamonadaceae</i>	<i>Delftia</i>	1
			<i>Oxalobacteraceae</i>	<i>Massilia</i>	1
<i>Gammaproteobacteria</i>	<i>Oceanospirillales</i>		<i>Halomonadaceae</i>	<i>Aidingimonas</i>	2
				<i>Halomonas</i>	10
				<i>Phytohalomonas</i>	1
		<i>Pseudomonadales</i>	<i>Pseudomonadaceae</i>	<i>Pseudomonas</i>	6
		<i>Xanthomonadales</i>	<i>Xanthomonadaceae</i>	<i>Lysobacter</i>	3
		<i>Chromatiales</i>	<i>Wenzhouxiangellaceae</i>	<i>Wenzhouxiangella</i>	1
		<i>Nevskiales</i>	<i>Steroidobacteraceae</i>	<i>Steroidobacter</i>	1
		<i>Cellvibrionales</i>	<i>Microbulbiferaceae</i>	<i>Microbulbifer</i>	1
		<i>Enterobacteriales</i>	<i>Enterobacteriaceae</i>	<i>Edaphovirga</i>	1
<i>Bacillota</i>	<i>Bacilli</i>	<i>Bacillales</i>	<i>Bacillaceae</i>	<i>Bacillus</i>	11
				<i>Oceanobacillus</i>	6
				<i>Gracilibacillus</i>	5
				<i>Virgibacillus</i>	2
				<i>Tenuibacillus</i>	2
				<i>Lentibacillus</i>	2
				<i>Halobacillus</i>	2
				<i>Salinibacillus</i>	1
				<i>Radiobacillus</i>	1
				<i>Aquibacillus</i>	1
				<i>Alkalicoccus</i>	1
				<i>Alkalibacillus</i>	1
				<i>Aidingibacillus</i>	1
				<i>Terribacillus</i>	1
			<i>Paenibacillaceae</i>	<i>Paenibacillus</i>	8
				<i>Saccharibacillus</i>	1

(待续)

(续表 1)

Phylum	Class	Order	Family	Genus	Genus number
<i>Bacteroidota</i>	<i>Cytophagia</i>	<i>Cytophagales</i>	<i>Hymenobacteraceae</i>	<i>Cohnella</i>	1
				<i>Planomicrobium</i>	1
				<i>Enterococcaceae</i>	1
				<i>Enterococcus</i>	1
				<i>Clostridiaceae</i>	1
				<i>Anaerophilus</i>	1
				<i>Alkaliphilus</i>	1
				<i>Pontibacter</i>	9
				<i>Hymenobacter</i>	2
				<i>Rufibacter</i>	1
<i>Flavobacteriia</i>	<i>Flavobacteriales</i>	<i>Flavobacteriaceae</i>	<i>Cyclobacteriaceae</i>	<i>Botryobacter</i>	1
				<i>Cyclobacterium</i>	2
				<i>Litoribacter</i>	1
				<i>Cesiribacteraceae</i>	1
				<i>Nafulsella</i>	1
				<i>Confluentibacter</i>	2
				<i>Flavobacterium</i>	8
				<i>Paucihalobacter</i>	1
				<i>Weeksellaceae</i>	2
				<i>Chryseobacterium</i>	2
<i>Chitinophagia</i>	<i>Chitinophagales</i>	<i>Chitinophagaceae</i>		<i>Epilithonimonas</i>	1
				<i>Algoriella</i>	1
				<i>Niastella</i>	1
				<i>Flavitalea</i>	1
				<i>Chitinophaga</i>	1
				<i>Parapedobacter</i>	1
				<i>Sphingobacterium</i>	5
				<i>Arcticibacter</i>	1
				<i>Balneolia</i>	1
				<i>Balneolales</i>	1
<i>Deinococcota</i>	<i>Bacteroidia</i>	<i>Bacteroidales</i>	<i>Balneolaceae</i>	<i>Rhodohalobacter</i>	1
				<i>Deinococci</i>	1
				<i>Deinococcaceae</i>	4
				<i>Deinococcus</i>	4
				<i>Natrialbales</i>	2
				<i>Natrialbaceae</i>	2
				<i>Natratbaculum</i>	2
				<i>Saliphagus</i>	1
				<i>Natronorubrum</i>	1
				<i>Natrinema</i>	1
<i>Euryarchaeota</i>	<i>Halobacteria</i>	<i>Haloferacales</i>	<i>Haloferacaceae</i>	<i>Haloterrigena</i>	1
				<i>Halopiger</i>	1
				<i>Halopelagius</i>	2
				<i>Haloprofundus</i>	1
				<i>Halorubraceae</i>	2
				<i>Halolamina</i>	2
				<i>Halobaculum</i>	1
				<i>Halalkalicoccus</i>	1
				<i>Haladaptatus</i>	1
				<i>Candida</i>	1
<i>Ascomycota</i>	<i>Saccharomycetes</i>	<i>Saccharomycetales</i>	<i>Debaryomycetaceae</i>	<i>Udeniomycetes</i>	1
<i>Basidiomycota</i>	<i>Tremellomycetes</i>	<i>Cystofilobasidiales</i>	<i>Mrakiaceae</i>		

Haloactinospora、*Haloglycomyces*; 具有嗜碱特性的 *Phytoactinopolyspor*、*Haloactinopolyspora*; 耐受辐射的 *Radiobacillus*、*Desertibacter*、*Frigidibacter*; 嗜热特性的 *Amphiplicatus*; 兼具耐盐、耐碱、耐高温的广古菌门中 *Haladaptatus*、*Natribaculum*。表明通过新的分离培养策略可以获取新疆地区极端环境中更丰富的特有微生物资源, 为更深入的研究提供材料。

2.3 新种资源分离培养方法与培养基

从环境中分离并获得微生物的纯培养物, 是研究和利用微生物的关键步骤。在研究环境样品微生物多样性及组成时, 为更大限度分离获得微生物资源, 需要依靠高效且有针对性的分离培养策略。从统计发表新疆微生物新种的文献中可以发现, 分离策略主要体现在样品预处理、培养基设计、化学抑制剂等方面。

2.3.1 预处理

预处理指通过物理或者化学方法对样品进行预先处理, 便于能够分离出特殊功能微生物菌种。一般而言, 沙漠环境具有太阳辐射强、干旱等特征, 有学者采用 Co 辐射处理沙样, 分离得到能够耐辐射的菌种资源^[39-42]。鉴于耐辐射菌往往具有较高的 DNA 修复和解除活性氧(reactive oxygen species, ROS)胁迫的能力, 可应用于预防或治疗 DNA 损伤药物的开发^[43-44]。酚类添加物被推荐用于分离土壤中稀有放线菌类群, 如将土壤 100 °C 干燥 15 min, 后用 1.5% 苯酚和 0.03% 葡萄糖酸氯己定来分离链霉菌种^[45]。对于植物内生菌的筛选, 常选择 70% 乙醇和 2.5% 次氯酸钠溶液对植物组织进行表面消毒^[46]。目前针对自然环境中的土壤样品, 采用梯度稀释涂布分离纯培养物仍为主流策略。为筛选特定功能菌株, 研究人员多以目标功能为导向, 在分离菌株前对土壤样品进行预处理。如以某种污染物作为唯一碳源, 逐步富集土壤样品中

能够耐受并降解该污染物的菌株^[47-48], 便于后续菌株分离。

2.3.2 培养基设计

针对样品来源特点(含根际)或分离目标不同而采用不同的培养基是关键策略。经统计发现, 当分离源为冰川冻土样品时, 因 PYG 培养基对冷杆菌属(*Cryobacterium*)和黄杆菌属(*Flavobacterium*)等类群有较好的选择性, 研究者多优先选择 PYG 培养基进行菌种分离。由于年蒸发量大, 艾丁湖的 Na⁺浓度高达 127.76 g/L^[49], 七角井盐湖的 Na⁺浓度达 107.79 g/L^[50]; 针对高盐环境的盐湖微生物资源, 2008 年唐蜀昆等设计了一种多盐纤维素酪蛋白培养基(cellulose-casein-multisalts, CCMS)^[7], 并在盐度、pH 方面做调整以模拟真实环境条件, 培养出大量耐盐、嗜盐细菌。例如, 调高 pH 值用于分离嗜碱耐碱菌种^[51-53], 分离自甘家湖自然保护区碱土的 *Nocardiopsis ganjahuensis* HBUM 20038^T 对 pH 耐受高达 13^[54]; 增加盐浓度以分离嗜盐耐盐菌种^[55-56], 分离自高盐土壤的 *Streptomonospora alba* YIM 90003^T 对盐耐受度高达 25% (质量体积分数)^[55]; 在分离植物内生菌时, 有些研究者还采用了在培养基当中添加一定量的植物组织液等策略来提高培养获得率^[22,57-58]。

2.3.3 添加化学抑制剂/促进剂

在分离放线菌时, 在培养基中添加适量的萘啶酸(nalidixic acid)、氨曲南(aztreonam)、放线菌酮(cycloheximide)、制霉菌素(nystatin)等物质抑制革兰氏阴性菌或真菌的生长^[59-62]; 添加复苏促进因子(resuscitation-promoting factors, Rpf)复苏分离出活的非可培养状态(viable but non-culturable, VBNC)的新种^[63], 说明加入 Rpf 是分离 VBNC 菌株的一种可行的方法。

传统的稀释平板涂布法适合将群落中生长占优势的菌株分离出来, 富集培养法是将样品

中的低丰度菌株富集后再分离的一种有效方法，可减少重复筛选已报道菌种的概率；由富营养组分转变为寡营养组分是不利于大多数微生物生长的特定选择性培养基，但可分离到大量具有特殊生理特性，如嗜盐、耐盐和嗜碱、耐碱的微生物。在培养时加入特定菌属的生长抑制剂，加速目标菌属的筛选。加入微量元素、促生长因子等也可起到让某类目标菌属更好生长的效果。利用 16S rRNA 基因高通量测序信息及样品环境特点设计分离培养基，同样被证实可以提高目标菌属的分离培养成功率。

2.4 新种微生物的应用

众所周知，放线菌是抗生素和生物活性物质的重要来源。近年来，耐药微生物的出现导致从常见链霉菌属中寻找新型药物制剂难度加剧。因此，从新疆地区特殊环境中寻找新链霉菌属菌种成为研究热点之一，如分离自胡杨根际土壤的 *Streptomyces indoligenes* TRM 43006^T 产生的吲哚衍生物，能够抑制表皮葡萄球菌细胞膜的形成^[64]；分离自喀纳斯森林土壤的 *Streptomyces kanasensis* ZX01^T 能产生抗植物病毒物质糖蛋白 GP-1^[45]，对烟草花叶病毒尤其有效；*Streptomyces luozhongensis*、*Streptomyces taklimakanensis* 表现出抗真菌、细菌活性的能力，表明该菌种具有作为生防菌的潜力^[65-66]，但其次生代谢产物仍有待进一步研究。微生物代谢的多样性赋予了微生物特有的功能，微生物可以降解环境中的有机化合物，如 *Streptomyces polyasparticus* TRM66268-LWL^T 菌种加速消减掩埋在土壤中的聚天冬氨酸(polyaspartic acid, PASP)^[47]；*Arthrobacter sulfonylureivorans* LAM7117^T 5 d 内对氯噬磺隆降解率为 85.6%^[67]；*Actinoplanes solisilvae* LAM7112^T 对盐酸四环素的降解率为 65%^[48]。这些具有特定功能的菌种在工业生产和环境治理方面均具有潜在的开发价值。

通过对我国新疆地区新种微生物资源的研究，挖掘到了许多具有地域特色的微生物，其中分离自高盐环境(如盐湖、盐土)耐盐、耐碱的菌种资源可以用于盐碱地的改良。分离自古海热泉的耐热菌对耐受的温度高达 65 °C^[68]，可以作为工业酶制剂、堆肥的菌种资源。沙漠环境中的放线菌纲中还存在一类对离子辐射具有超强抵抗能力的耐辐射动球菌，该属很有可能存在着非同源末端连接(non-homologous end joining, NHEJ)系统高效修复双链断裂(double-strand breaks, DSB)这类 DNA 常见损伤。分离自冰川冻土环境的嗜冷、耐冷菌株，也可作为低温酶制剂等的来源。所有这些长期生活在极端生境中的菌种，它们形成了独特的生理学特性和适应机制，其生物膜的结构、酶的分子结构、基因的表达和调控都值得进行深入的研究。在工业生物技术领域，生长在特殊环境中的极端微生物，如嗜盐菌、嗜酸菌、嗜碱菌、嗜冷菌、嗜热菌等可作为底盘细胞，由于其生长在苛刻的环境中，可以避免杂菌污染对发酵培养的干扰^[69]。综上所述，在新种微生物应用方面已得到了部分可以产生生物活性物质和可用于环境治理的新种资源。

3 总结与展望

国内外对新疆地区微生物新种资源的研究很广泛，包括特殊生境(盐生植物根际、冰川、沙漠)和极端环境(高盐、高碱、强辐射)的细菌、古菌及真菌。本文介绍了这些新种的生境特点、分离方法、研究单位、功能特性等，总结了这个发掘过程的发展趋势、分离培养基类型及分离方法，阐述了新疆地区不同环境中孕育着多样性极高的菌种资源。

目前，微生物在人们生产、生活各个方面 的应用十分广泛。在工业上，细菌的次级代谢产物

可以产生大量具有活性的化合物、抗菌物质用于开发生产。Zhou 等以 *Streptomyces kanasensis* ZX01 发酵产物中的糖蛋白 GP-1 为有效成分研制出一种新型的抗植物病毒制剂, 室内盆栽与田间小区试验均表明 0.5% 糖蛋白 GP-1 水剂在防治烟草花叶病毒病方面表现出 45%–75% 的持效性^[70]。在环境污染的治理中, 具有降解基因或降解机理的细菌可用于生态修复。在盐碱地的改良中, 耐盐碱的细菌通过代谢活动改善土壤状况, 降低盐碱浓度和含量。对于我国新疆特殊环境地区新种资源的研究, 尚需关注以下几个方面: (1) 产生物活性物质菌属(放线菌)的研究与利用; (2) 新疆非豆科植物共生固氮资源的挖掘与鉴定; (3) 能够产生特殊代谢产物菌种的深入研究与开发利用。因此, 对我国新疆特殊生境新种资源的挖掘是具有极高的科学价值、生态价值和经济价值。

传统的新种多相分类鉴定包含形态观察、生理生化鉴定、化学分类及分子分类, 现在又发展到基因组水平鉴定。如何进一步巩固、创新, 开发出更快速便捷的鉴定技术、方法、扩充相对应的数据库, 收录相关的分离方法、培养基、样品处理方式, 以及基因层面的功能和生态层面的功能特性成为难题。近年来, 运用宏基因组技术对未培养类群进行生理代谢功能(如碳、氮源利用情况)预测的研究大量涌现, 可据此设计一些稀有类群的分离方案, 用以分离指定菌株。此外, 在线网站或科研工具(如 KOMODO)也能够帮助研究者有针对性地设计分离培养基和培养条件^[71]。通过对我国新疆地区各生态环境微生物菌种资源的开发研究, 对于理解特殊生态系统功能、开发利用极端环境微生物资源具有重要的意义; 更为重要的是做好分离培养方法的开发和创新, 借此丰富菌种资源库, 加大对特殊功能菌种资源的研究与开

发, 可为人类的生活、生产提供更加充足的资源保障。

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